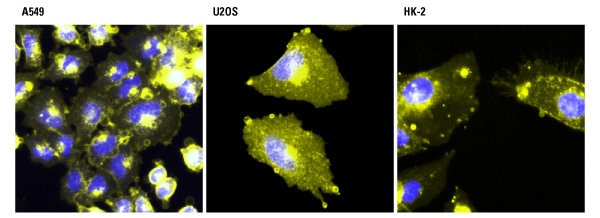
# **Metabolic Labeling Strategies**

Metabolic labeling refers to methods in which the endogenous synthesis and modification machinery of living cells is used to incorporate detection or affinity tags into biomolecules. Typically, this is accomplished by culturing cells or organisms in media in which a specific natural molecular building block (e.g., amino acid, nucleotide, carbohydrate) has been replaced with a tagged chemical analog. Cells use the chemical analog instead of the natural biomolecule to synthesize or modify proteins, nucleic acids, etc. An alternative method of metabolic labeling involves genetically engineering the cellular machinery to accomplish site-specific biomolecule labeling or to incorporate chemical analogs that are not tolerated by endogenous synthesis and modification machinery. Metabolic labeling is a powerful strategy because it is simple to perform and enables measurement of metabolic rates and detection of biologically relevant interactions *in vivo*.

Radioactive metabolic labeling

Radioactive analogs are the most commonly used compounds for metabolic labeling of cells and organisms. Since radiolabeled isotopes can be substituted in biomolecule monomers without any changes to the chemical structure, they are readily incorporated *in vivo*. Radiolabeled macromolecules are also easily detected by sensitive radiometric techniques such as liquid scintillation counting or positron emission tomography (PET) scanning. Examples of radioactive tracers and applications include 3H thymidine uptake for cell proliferation assays, 35S methionine labeling for protein synthesis determination, 32P orthophosphate labeling for *in vivo* kinase assays, and 14C-labeled D-glucose update for determination of cellular metabolism rates. Although radioactive isotopes are easily detected and relatively inexpensive, there are some disadvantages including safety hazards, generation of radioactive waste, toxicity to organisms, and radioactive decay leading to loss of signal over time.

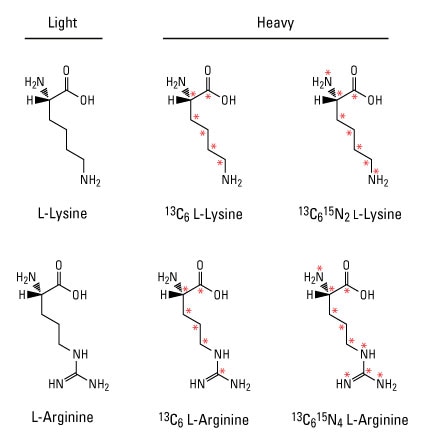
This representative experiment shows how azide-reactive Thermo Scientific DyLight dyes may be used to label cultured cells.



**Detection of metabolic labeling with azido-sugars in different cell types using azide-reactive DyLight dyes**. A549, U2OS and HK-2 cells were incubated with 40 µM azido-acetylmannosamine in cell culture media for 72 hours and then incubated with 100 µM of Thermo Scientific DyLight 550-Phosphine (yellow). The cells were washed, fixed with 4% paraformaldehyde and counterstained with Thermo Scientific Pierce Hoechst 33342 (blue).

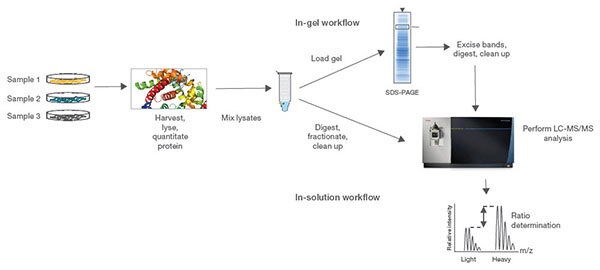
Stable isotope labeling using SILAC

In contrast to radioactive metabolic labeling, stable isotope labeling uses isotopic analogs of biomolecule monomers which are not radioactive. Like radioisotopes, stable isotopes can be incorporated into biomolecule monomers without any changes to the chemical structure. Common stable isotopes used for metabolic labeling include 2H, 15N, 13C and 18O. Since they are not radioactive, these heavy stable isotopes are detected by their increased mass compared to natural light isotopes using mass spectrometry (MS).



**Example light and heavy amino acids for SILAC.** Red asterisks indicate positions of stable isotopes (13C and 15N). Incorporation of respective light and heavy amino acids into proteins by cells in various treatment conditions can be measured by mass spectrometry.

Stable isotope labeling by amino acids in cell culture (SILAC) is the most common approach for metabolic labeling using stable isotopes. In this method, cells are cultured in special growth medium that contains either light or heavy amino acids such as 13C6-lysine and 13C6-arginine. After labeling, light and heavy samples are combined to measure differences in relative protein expression or post-translational modifications by MS. This powerful quantitative proteomic technique is particularly useful to detect relatively small changes in protein levels and can be multiplexed to measure up to three different experimental conditions in a single MS analysis.

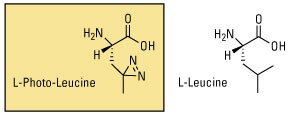


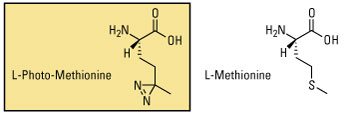
**Procedure summary for MS experiments using SILAC reagents.** Normalized protein extracts isolated from cells are combined, reduced, alkylated and digested overnight. For the in-gel workflow, samples are run on an SDS-PAGE gel, excised, digested and cleaned up; for the in-solution workflow, samples are digested, fractionated and cleaned up. Samples are then analyzed by high-resolution Thermo Scientific Orbitrap LC-MS/MS.

Photoreactive amino acids

Photoreactive amino acids are chemical analogs of amino acids that enable metabolic labeling and *in vivo* crosslinking. These amino acids contain a small diazirine functional group which is stable under normal lab environments but can be photoactivated with long-wave UV light (330 to 370 nm). Upon UV activation, the diazirine group is liberated to generate reactive carbene intermediates, which covalently bond to any amino acid side chain or peptide backbone in close proximity.

Photo-L-Leucine and Photo-L-Methionine are two examples of photoreactive amino acids designed for metabolic labeling of proteins. An advantage of using these amino acids for studying protein–protein interactions is the ability to crosslink proteins at their protein interaction interfaces. However, since these amino acids are not identical in structure to their natural counterparts, photoreactive amino acids typically have lower incorporation rates than isotopic analogs and are not well tolerated by cells or complex organisms for long growth periods.





**Photoreactive amino acids.**When grown in media devoid of the natural amino acids, cells will incorporate the photoreactive analogs. Upon subsequent exposure to UV light, the diazirine group will crosslink to any molecules within its reach (such as an interacting polypeptide chain).

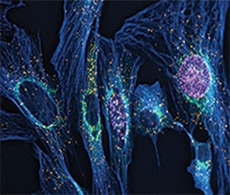
Bioorthogonal labeling reagents

Bioorthogonal labeling reagents are metabolic labeling compounds whose functional groups can be chemically reacted without interfering with biological processes. Bioorthogonal functional groups that have been used as the basis for metabolic labeling compounds include azides, alkynes, aldehydes and ketones. Unlike radioactive or stable isotope analogs, bioorthogonal analogs are not detected directly but rather reacted with another detection or affinity compound through a chemoselective ligation reaction.

One chemoselective ligation reaction is a copper-catalyzed Huisgen cycloaddition between azides and alkynes, commonly referred to as Click chemistry. Either azides or alkynes can be used as the functional group for metabolic labeling due to their small size and compatibility with biological systems. However, traditional Click chemistry reactions use copper as a catalyst, which is toxic to living organisms. Therefore, copper-free Click probes using larger cycloalkynes have been developed as an alternative method for ligation.

The Staudinger ligation is another chemoselective chemistry which involves the reaction of azido bioorthogonal probes with phosphine compounds. Similar to Click chemistry, the ligation reaction is highly specific and can be performed in aqueous environments at physiological pH. However, unlike traditional Click chemical reactions, Staudinger ligations do not require copper to be reactive. Although this increases the biocompatibility of Staudinger ligations, these reactions also tend to be slower than Click reactions due to the absence of a catalyst.

Click chemistry is the detection method of choice for samples that would be compromised by direct labeling or antibody-based secondary detection techniques. The click label is small enough to penetrate complex samples easily, and the selectivity and stability of the Click reaction provides high sensitivity and low background signal. This gentle sample treatment together with the biocompatible Invitrogen Click-iT Plus reaction means that detection can be multiplexed with expressed proteins such as GFP, protein labels such as R-PE, and a wide range of organic fluorophores.

[](https://www.thermofisher.com/eg/en/home/life-science/protein-biology/protein-labeling-crosslinking/protein-labeling/metabolic-labeling-reagents-proteins.html)

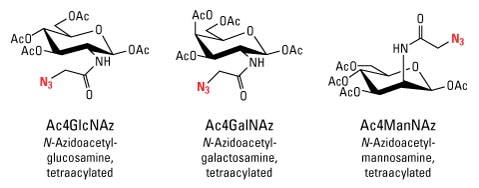
**Detecting of proliferating HeLa cells.** Measuring DNA synthesis is the most precise way to detect changes in cell proliferation. Image-based proliferation assays generate spatial and temporal results that cannot be detected with other methods. In this example, proliferation of HeLa cells was detected using [Invitrogen Click-iT Plus EdU Alexa Fluor 488 Imaging Kit.](https://www.thermofisher.com/order/catalog/product/C10637)

**Watch this video to learn more about Click chemistry**

Play Video

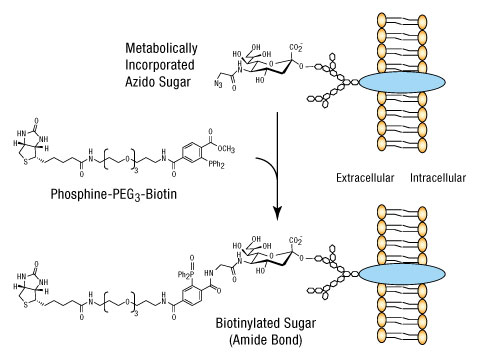
**Bioorthogonal probes**

Numerous bioorthogonal probes have been developed for metabolic labeling, including azidohomoanaline for labeling newly synthesized proteins, 5-ethynyl-2'-deoxyuridine (EdU) for labeling nucleic acids, azido palmitic acid for labeling inter-membrane proteins, and azido sugars for labeling glycoproteins. Azido sugar reagents were one of the first bioorthogonal probes used for metabolic labeling. After incorporation of the azido sugar into N-linked glycans, they can be chemoselectively targeted with alkyne- or phosphine-derivatized compounds for detection or affinity purification.



**Azido sugars for metabolic labeling of glycoproteins.**

Due to the wide variety of detection and affinity compounds available for ligation ranging from biotins to fluorescent dyes to mass tags, bioorthogonal labeling is one of the most versatile methods for metabolically labeling biomolecules. The illustration provides an example of how biotinylated sugar moieties may be used for *in vivo* labeling.



**Example strategy for *in vivo* metabolic labeling with biotin using azide–phosphine reagents.** When azido–sugar derivatives are supplied to living cells, they are incorporated into glycoproteins by endogenous post-translational modification mechanisms.The azide-tagged molecules can then be selectively labeled or conjugated to phosphine-activated molecules, in this case, a derivative of biotin. If the azido sugars were supplied to cells being studied for response to a particular treatment, the biotin affinity tag could be used to purify and analyze differences in glycosylation resulting from the treatment regime.

Site-specific metabolic labeling

Metabolic labeling of biomolecules distributes the label throughout the molecule based on the biomolecule composition. For example, culturing cells with 13C6-lysine results in the incorporation (labeling) of all lysine-containing proteins synthesized during the experiment. Because most metabolic labeling strategies use chemical analogs which are structurally identical or very similar to natural monomers, it is usually not possible to control the specific site of incorporation or the number of labels added. However, site-specific metabolic labeling is possible when metabolic probes are designed and used in combination with genetically altered organisms or cell-free *in vitro* translation systems.

One method for site-specific incorporation takes advantage of redundant stop codons present in the genetic code, which can be used to incorporate amino acid analogs using specifically engineered tRNAs.  In addition to allowing for site-specific incorporation, this method allows for labeling with chemical analogs such as biotinylated or fluorescent dye-labeled amino acids which are not tolerated by endogenous synthesis and modification machinery. This method also limits the incorporation of the metabolic label to the expressed, genetically modified proteins.

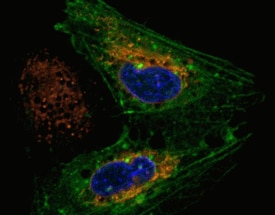
Fluorescent probes used in live cells

Fluorescent molecules, also called fluorophores or simply fluors, respond directly and distinctly to light and produce a detectable signal. Unlike enzymes or biotin, fluorescent labels do not require additional reagents for detection. This feature makes fluorophores extremely versatile and the new standard in detecting protein location and activation, identifying protein complex formation and conformational changes, and monitoring biological processes *in vivo*.

The vast selection of fluorophores today provides greater flexibility, variation and fluorophore performance for research applications than ever before. Fluorophores can be divided into three general groups, and each group of probes has distinct characteristics. These groups are as follows:

* Organic dyes—FITC, TRITC, DyLight fluors
* Biological fluorophores—Green fluorescent protein (GFP), R-Phycoerythrin
* Quantum dots

Detection of fluorescent probes requires specialized equipment, including an excitation light source, filter set and a detector, that are found in fluorescence microscopes, fluorescence plate-readers, flow cytometers and cell sorters. This equipment enables the detection of very small levels of fluorescent probes, allowing highly sensitive assays to be generated. Thermo Fisher Scientific offers a wide variety of fluorescent probes that allow researchers to detect variations in cellular physiology, including changes in cell proliferation, cell death and other changes associated with disease states or microbial invasion of the cellular host.



**Live cell imaging.** Detection of oxidative stress in the human osteosarcoma (U20S) cell line.

Live Cell Imaging eBook

Published by Wiley, this free eBook provides essential knowledge and provides a short, practical guide of the background and basics of live cell imaging. Content includes:

* Basic history of live cell imaging
* Case studies of live cell imaging applications
* Common problems and solutions with live cell imaging